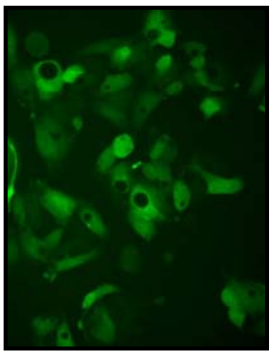
REF: P20111

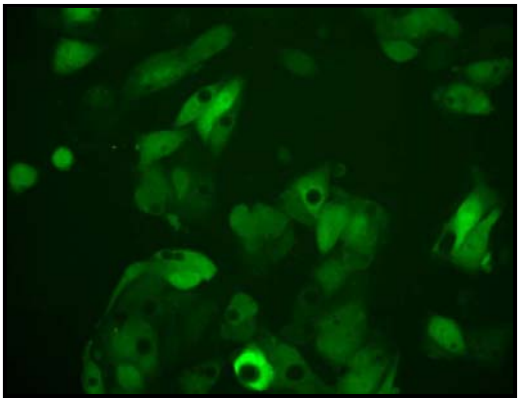
LINTERNA™ CELL LINES

GREEN FLUORESCENT HEP-G2 CELLS



Product Name:	LINTERNA™ – HEP-G2 Cell line
Catalog Number:	P20111
Cell Line:	Hepatocellular carcinoma
Fluorescent Protein:	tGFP
Format:	3 x 10^6 cells in Cryopreserved vials
Storage:	Liquid Nitrogen

A novel green fluorescent HEP-G2 cell line has been developed through stable transfection with Evrogen TurboGFP. This cell line expresses green fluorescent protein gene sequences as free cytoplasmatic proteins.



tGFP-HEP-G2 Cell line is stably-transfected clonal cell line that is ready to use in cell-based assay applications. This stably transfected clonal cell line provides consistent levels of expression, which helps to simplify the interpretation of results. This cell line is intended to be used as "in vitro" model for research studies.

About HEP-G2

The Hep G2 cell line has been isolated from a liver biopsy of a Caucasian male aged 15 years in 1975, with a well differentiated hepatocellular carcinoma. They are epithelial-like cells, adherent and growing as monolayers and in small aggregates. Cells reportedly produce a variety of proteins: alpha-fetoprotein, albumin, alpha2-macroglobulin, alpha1-antitrypsin, transferrin, alpha1-antichymotrypsin, haptoglobin, ceruloplasmin, plasminogen, complement (C3, C4), C3 activator, fibrinogen, alpha1-acid glycoprotein, alpha2-HS glycoprotein, ß-lipoprotein, retinol binding protein, 3-hydroxy-3-methylglutaryl-CoA reductase and hepatic triglyceride lipase activities. There is no evidence of a Hepatitis B virus genome in this cell line.

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About TurboGFP

tGFP is an improved variant of the green fluorescent protein CopGFP cloned from copepoda Pontellina plumata (Arthropoda; Crustacea; Maxillopoda; Copepoda). It possesses bright green fluorescence (excitation/ emission max = 482/ 502 nm) that is visible earlier than fluorescence of other green fluorescent proteins. tGFP is mainly intended for applications where fast appearance of bright fluorescence is crucial. It is specially recommended for cell and organelle labeling and tracking the promoter activity.

Quality Control

All cells are performance assayed and test negative for mycoplasma, bacteria, yeast and fungi. Cell viability, morphology and proliferative capacity are measured after recovery from cryopreservation. Innoprot guarantees stable expression for many generations and provides support for cell culture and visualization.

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CELL CULTURE INSTRUCTIONS

A. Complete Growth medium

DMEM (D6429 from Sigma -Aldrich)
10% FBS
G418: 500 µg/ml

B. Set up culture after receiving

1. Decontaminate the external surfaces of medium and medium supplements with 70% ethanol.

2. Prepare coated flask (T-75 flask is recommended). Add 9 ml of DMEM and 1 ml of FBS (without selection antibiotic). Leave the flask in incubator minimum one hour at 37ºC incubator.

3. Place the vial in a 37ºC waterbath, hold and rotate the vial gently until the contents are completely thawed. Remove the vial from the waterbath immediately, wipe it dry and rinse the vial with 70% ethanol. Remove the cap, being careful not to touch the interior threads with fingers.

4. Dispense the contents of the vial using 1 ml eppendorf pipette and gently resuspend the contents of the vial in T75 flask containing pre-warmed complete growth media.

6. For best result, do not disturb the culture for 16 hours after the culture has been initiated. Change the growth medium (including the selection antibiotic) the next day to remove the DMSO and unattached cells.

Maintenance of Culture:

1. Change the medium to fresh supplemented medium the next morning after establishing a culture from cryopreserved cells. For subsequent subcultures, change medium 48 hours after establishing the subculture.

2. Once the culture reaches 50% confluence, change medium every day until the culture is approximately 80% confluent.

3. Subculture the cells when they are over 90% confluent.

4. Incubate cells with 1 ml of trypsin/EDTA solution (in the case of T-75 flask) until 80% of cells are rounded up (monitored with microscope). Add 1ml of trypsin neutralization solution to the digestion immediately and gently rock the culture vessel.

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